

## Methods in Quaternary Ecology #4. Freshwater Sponges

Frederick W. Harrison  
Department of Biology  
Western Carolina University  
Cullowhee, North Carolina 28723

### Summary

In recent years, there has been an increased awareness of the value of fossilized remains of freshwater sponges in Quaternary paleoecological reconstructions. Because the taxonomy and ecology of extant forms have been clarified, and because a well-defined fossil record exists that exhibits continuity

with extant species, the stage is set for widespread utilization of freshwater sponge remains in Quaternary environmental reconstructions. This paper presents an introduction to the freshwater sponges, techniques for their utilization in Quaternary paleoecological studies, and summarizes some of the most recent work performed in North America.

### Introduction

Despite the astonishment of many individuals, sponges do exist in most freshwater habitats and in fact, have existed for at least 100 million years. We are familiar with marine sponges. Their often brilliant hues attract attention and even if most sponges purchased from the local merchant consist of a non-natural plastic, many of us still remember the day when a "bath sponge" was non-synthetic. But what of these freshwater sponges? What do they look like (Figures 1 and 2), where can one find them, are they *truly* sponges, and how are they important in Quaternary ecology?

### Freshwater Sponge Biology: An overview

Freshwater sponges are members of the great poriferan class Demospongiae, a taxonomic grouping that contains most species

of marine sponges. The majority of freshwater sponges are members of the family Spongillidae, a widely distributed group characterized by a siliceous spicular skeleton and by asexual reproductive structures, gemmules. Fortunately for the paleoecologist, both these diagnostic sponge parts are found as fossils in lacustrine and peat deposits.

As with all sponges, the physiology of freshwater forms is organized around a system of canals that courses through the body of the animal (Figure 3). Water is driven through the canals by clusters of flagellated cells, choanocytes; thus, the animal is in intimate contact with the aquatic environment. Because of this degree of dependency, the sponges are delicate monitors of water quality, degenerating or dying in the presence of adverse conditions. But under favourable circumstances, the sponges utilize their aquatic environment, filtering their bacterial food from it, assimilating dissolved oxygen, and releasing waste products into the excurrent flow of water in their canal systems.

In terms of most criteria, the freshwater sponges are organized and exist upon the same ancient organizational and physiological patterns as most marine sponges. They differ from most marine sponges in their mode of reproduction. While they do reproduce sexually, freshwater sponges also survive the harsh conditions of northern winters or the warm waters of southern summers by entering an asexual phase of resistant gemmules.

The habit of gemmule formation apparently evolved as a response to unstable environmental conditions. This is seen in a few marine species, particularly those found in harsh, fluctuating intertidal environments.

Seasonal environmental changes trigger the release of endogenous stimuli, perhaps cyclic nucleotides, in the body of the sponge. The amoeboid cells of the interior of the sponge migrate and mass in spherical aggregates. As gemmule development proceeds, cells at the aggregate periphery assume a secretory

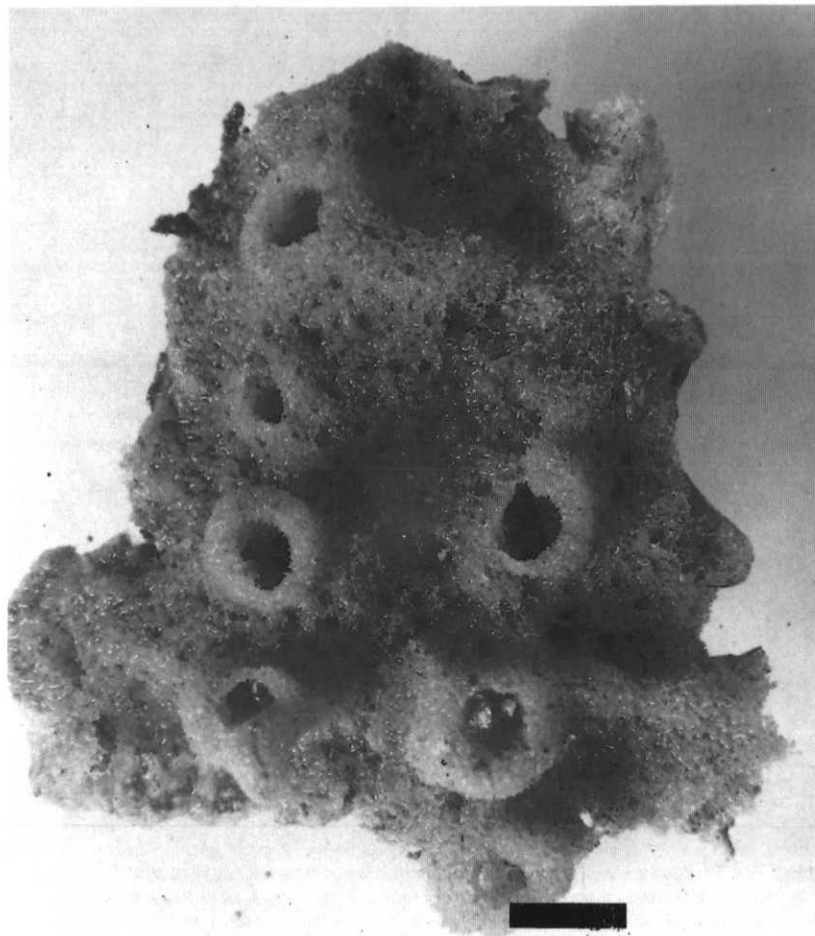


Figure 1 Modern freshwater sponge, *Eunapius carteri*. The large openings are oscules. Scale bar = 1 cm.

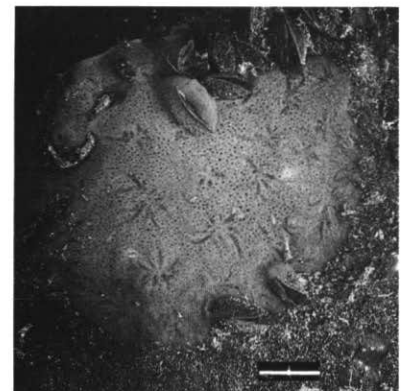


Figure 2 Modern freshwater sponge, *Eunapius fragilis*. Scale bar equals 1 cm.

function, depositing a shell of spongin, a form of collagen. Cells in the aggregate interior accumulate yolk in lens-shaped platelets which crowd the cytoplasm of these reproductive cells. However, the gemmules with their protective shells of spongin survive and await the coming of more favourable conditions. With the return of warmer, or cooler, water temperatures, a burst of germinative activity sends the yolk cells of the gemmule migrating out over its surface, spreading upon the substrate, and differentiating into a young sponge. There the sponge grows, forming an encrusting green, tan, or darkly coloured mass on emergent vegetation, upon floating logs, or in the folds and ridges of submerged stones.

The preceding paragraphs offer a brief overview of limited aspects of freshwater sponge biology. The biology of sponges has been reviewed in detail in recent publications. Those seeking further information about these animals should begin with the excellent book by Simpson (1984), *The Cell Biology of Sponges*.

Eventually, siliceous spicules comprising the skeleton of the sponge, and the hard, resistant shells of gemmules become incorporated in the sediment. Although spicules and gemmules which are easily recognizable and diagnostic are preserved in the fossil record, they have not played a significant role in paleoecological reconstructions. Their use has been overlooked by paleoecologists due

to a dearth of knowledge of the ecology of extant freshwater sponge species and long-standing confusion about the taxonomy of the group. Recent advances in both areas of study finally allow the utilization of fossilized sponge remains in paleoecology.

#### Freshwater sponges in paleoecology:

##### The historical perspective

For freshwater sponges to be useful in paleoecological studies, it is necessary for three factors to exist. First, it is necessary to have a well-defined fossil record that exhibits continuity with extant species. Second, an orderly systematic framework must be available so that the non-specialist on sponges may identify his materials. Finally, the environmental preferences, including physicochemical parameters, of species must be understood. Great effort has been made within the past two decades to meet these three conditions, thus setting the stage for widespread use of this group by paleoecologists.

Much of our understanding of freshwater sponge systematics dates from the monumental study by A.A. Racek (Penney and Racek, 1968). Racek, in completing the study only just initiated before the death of Professor Penney, brought order out of the chaos of spongillid systematics by recognizing global evolutionary patterns among spongillid species. He demonstrated the artificial nature of the two spongillid sub-

families Spongillinae and Meyeninae, erecting a new genus, *Radiospongilla*, for those sponges showing morphological similarities to the two subfamilies. Although lacking fossil evidence, Racek proposed that the two artificial subfamilies had diverged from a common ancestral stock, a stock that had also given rise to the genus *Radiospongilla*. The fossil spongillid, *Paleospongilla chubutensis*, recovered from Cretaceous strata, exhibits spicular characters combining those found in the two subfamilies, thus suggesting the derivation of both subfamilies and appearing to be the most direct ancestral form of the genus *Radiospongilla* (Racek and Harrison, 1975). With the clarification of spongillid systematics, recent taxonomic treatments and keys (Pennak, 1978) have followed the Penney and Racek (1968) nomenclature. Therefore, as spicules and gemmules (Figures 4 to 8), the same elements that are preserved as fossils, are diagnostic criteria for fossil sponge identification, specialists may identify sponges both extant and fossil with the same level of confidence. Recently, increasing work, particularly on North American species has allowed for a fuller understanding of sponge ecology. The ecology of individual species of North American freshwater sponges is presented in a series of papers by Harrison (1974, 1977, 1979), Harrison *et al.* (1977), Harrison and Harrison (1977, 1979), Jewell (1935, 1939), Old (1932), and Poirrier (1969, 1974, 1977).



**Figure 3** Diagram of the anatomy of a freshwater sponge "Knobbed" arrows indicate incurrent flow of water. Arrows without "knobs" indicate excurrent flow. A superior epithelium consisting of a layer of fusiform external pinacocytes and an inner layer of endopinacocytes (black cells) is pierced by pores (p). Environmental water passes through the surface epithelium, into a sub-epithelial space (atrium, at), and into the loosely organized mesohyl (m). Water is both drawn into and expelled from choanocyte chambers (c) by the beating of choanocyte (collar cell) flagella. The excurrent system terminates in oscular chimneys (o). Water exits the sponge by oscules, openings at the top of the chimney. Ameboid cells (a) migrate through the mesohyl. Skeletal elements, spicules (sp), and the basopinacoderm (b) attachment surface respectively, are bound into fascicles or attach through the matrix material, spongin (s)

Consequently, the three necessary criteria have been met. A fossil record with direct relationships to extant forms dates from Cretaceous time. Freshwater sponge species exist within a well-defined systematic framework and, important for limnologic analysis, the presence of particular species may be useful in characterizing and defining local aquatic environments.

Therefore, building upon earlier European studies (Frey, 1964) and upon pioneering studies by Racek (1966, 1970, 1974) of deposits from Europe, Jordan, and Central America, there has been increasing interest in freshwater deposits particularly in North America (Hall and Hermann, 1980; Harrison *et al.*, 1979; Turner, 1985; Harrison and Warner, 1986).

#### Field Sampling and Laboratory Extraction Techniques

In the field, lake and peatland cores can be collected with standard coring equipment. For spicule analysis, subsamples are boiled in 1:1 mixture of concentrated nitric and sulphuric acids until a dry white dust remains. Following washing, this residue is retained for microscopic examination. In addition, specific volumes (usually 20 cm<sup>3</sup>) of fresh sediment are washed on a 250 µm mesh sieve, and the remaining residue is examined for macroscopic gemmules under a dissecting microscope.

Identification of freshwater sponge remains relies upon analysis of spicules and gemmules. There are three spicule types in

freshwater sponges: megascleres, gemmoscleres, and microscleres. The largest, megascleres, are skeletal spicules which form a supportive framework for the delicate sponge tissues. Megascleres are of limited taxonomic value although they are the predominant spicule type in the sponge. Gemmoscleres which are components of the gemmule coat form the armature of gemmules. Their morphology and placement in the gemmule coat are of considerable diagnostic value in taxonomy. Microscleres, the small "dermal" spicules, are not found in all species. However, their presence or absence and their morphology are of considerable value in taxonomic determinations. A taxonomic key to North American freshwater sponges is presented in Pennak (1978). The original Pennak and Racek (1968) monograph is valuable because of the clarity of illustrations. The scanning electron microscope promises to be of exceptional value in taxonomic analysis, but at present, has been rarely used (Harrison, 1981).

#### Quaternary Environmental Reconstructions

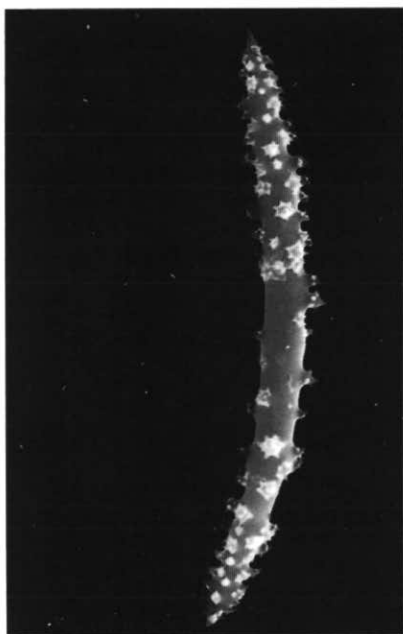
Although there have been few studies utilizing freshwater sponge remains, the most recent studies involve examination of lake sediments from Canada and from Kentucky, USA. Analyses of fossil spicules and whole gemmules of freshwater sponges were performed on early Holocene sediments collected from Serendipity Bog, a bog lake on eastern Graham Island, Queen Charlotte Islands, British Columbia (Harrison and Warner, 1986). Their study established the essential criterion of corroboration of data

obtained from analyses of freshwater sponge remains with data derived independently from other fossilized components, e.g., pollen and plant macrofossils.

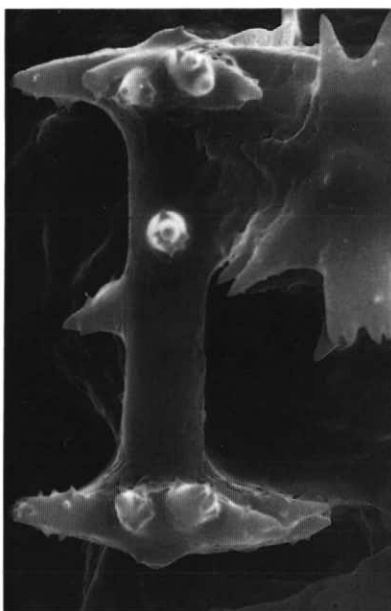
Examination of the spicular components in limnic sediments from the Serendipity Bog site revealed a flourishing freshwater sponge population between about 9400 and 8500 years B.P., composed of *Spongilla lacustris* and a member of the genus *Anheteromeyenia*. Of the two species, apparently, *Anheteromeyenia* was the community dominant throughout this period. Two peaks of fossil spicules, one at 3.15-3.10 m and another at 2.95-2.90 m depth, indicate that there were two major periods exceptionally favourable to the growth of sponges. Of the species represented in the Serendipity core, members of the genus *Anheteromeyenia* generally are more specific in their environmental requirements than is the cosmopolitan *S. lacustris*.

Sponges of the genus *Anheteromeyenia* prefer waters of moderate to low alkalinities (known range, 2-80 mg CaCO<sub>3</sub>·L<sup>-1</sup>; Harrison, 1974, 1977) that have high specific conductances (known range, 37-750 µmhos; Old, 1932; Jewell, 1939; Poirrier, 1969; Harrison, 1974, 1977). They prefer waters that are slightly acidic, but will grow well in circum-neutral to slightly basic habitats (known range, pH 4.2-8.5; Harrison, 1974, 1977).

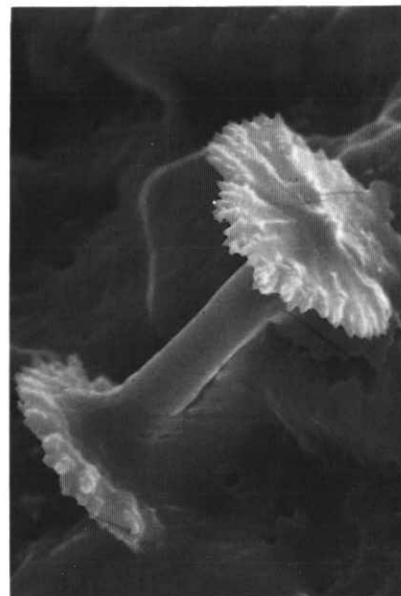
Extant *S. lacustris* exhibits broader habitat tolerances, although Poirrier (1969) notes that the species is common in acidic water low in bicarbonate; that it exists through a



**Figure 4** Scanning electron micrograph of a microsclere of *Spongilla lacustris*. Although this spicule type is not found in all freshwater sponge species, when present, the microscleres are of considerable diagnostic value. Magnification 2400 ×.



**Figure 5** Scanning electron micrograph of a birotulate gemmosclere of *Ephydatia fluviatilis*. The rotule of another spicule is seen in the upper right of the figure. Magnification 2000 ×.



**Figure 6** Scanning electron micrograph of a gemmosclere of *Anheteromeyenia ryderi*. Compare rotule morphology with that of the *E. fluviatilis* gemmosclere seen in Figure 5. This spicule is partly embedded in the spongin coat of the gemmule. Magnification 1800 ×.

wide range of pH; and that it shows a slight tolerance to basic waters of higher specific conductances, silicon concentrations, and colour (*i.e.*, dissolved organics) (known ranges, total hardness as  $\text{CaCO}_3$ , 28.8-250.0  $\text{mg} \cdot \text{L}^{-1}$ ; pH 5.3-9.0; specific conductance, 9.4-470  $\mu\text{mhos}$ ; silicon 0-20.5 ppm; colour 0-202; Harrison, 1974). Neither of these species is capable of withstanding high levels of turbidity, since suspended silt and clay particles tend to clog the canal system of the sponge's body.

Spicule development in *S. lacustris* and in other freshwater sponges is related directly to silicon levels (Jewell, 1939). Conspicuously abnormal (*i.e.*, underdeveloped) skeletal elements from *S. lacustris* were not observed in the fossil sequence, which suggested moderate to high levels of silicon in the water, an interpretation proffered in consideration of the quartzo-feldspathic sand deposits that lie beneath the Serendipity basin.

Available ecological information confirms that *S. lacustris* prefers habitats with high light intensity. Modern colonies of *S. lacustris* which were attached to small crystalline rocks, associated with *Isoetes echinospora* on the shore of Hippa Lake, west of Graham Island were observed in water about 15 cm deep. Spicules of *S. lacustris* appeared to be more abundant near the base of the Serendipity core, and progressively declined upward. Paleobotanical data indicated the presence of submerged and floating-leaved aquatic macrophytes (Warner, 1984), which

may suggest higher light intensities during earlier phases of development of the Serendipity basin. Subsequent macrophytic growth and progression to a fen community might have increased shading, and hence may have contributed to a reduction in the population of *S. lacustris*.

Members of the *Anheteromeyenia* group express variable responses to light. Perhaps the fossil remains of *Anheteromeyenia* were in fact those of *A. ryderi*, a species known to prefer shaded habitats. This factor may explain its better representation in the fossil sequence, inasmuch as it would be able to survive the shading effect of associated prolific macrophytic growth.

The presence of fossil materials from the submerged macrophyte *Potamogeton filiformis* corroborated limnological tolerances indicated by the freshwater sponge fauna. Like *S. lacustris* and *Anheteromeyenia* sp., *P. filiformis* occurs in circum-neutral to basic waters with pH usually above 5.9, and is indifferent to alkalinity of the water and specific conductance. However, *P. filiformis* almost always is confined to mineral substrates and to water from between 0.5 and 2.0 m deep (Warner, 1984).

Therefore, both the sponge fauna and the submerged aquatic flora as reconstructed from fossil evidence reflect that the early Serendipity basin was shallow. This basin contained standing or slow-moving waters that exhibited circum-neutral to slightly basic pH with moderate to low carbonate alkalinity and high specific conductance, silicon, and dissolved organic content.

The site of a second study utilizing fossil remains of freshwater sponges is Jackson Pond, a sink-hole lake located in the north-central region of Kentucky. The pond lies in

the northern portion of the Interior Low Plateaus and within 190 km of the full-glacial ice margin (Wilkins, 1985; Delcourt *et al.*, 1986). Jackson Pond sediments have been radiocarbon-dated from 20,000 years B.P. to the present (Wilkins, 1985).

Pollen records from Jackson Pond (Wilkins, 1985) indicate that the full-glacial (20,000 to 17,000 years B.P.) and late-glacial (17,000 to 11,500 years B.P.) intervals in central Kentucky were periods of cool, wet climate with high sedimentation rates in the pond, indicative of sheetwash erosion from the nearby uplands. These high sedimentation rates were associated with conditions of high levels of soil moisture (Delcourt, 1985; Wilkins, 1985). During the full-glacial interval, the region was occupied by closed boreal forest dominated by spruce with jack pine as a sub-dominant. After 17,000 years B.P., a period of open boreal woodlands characterized by both tree populations, but with an increase in herbaceous plants, persisted until the Holocene transition.

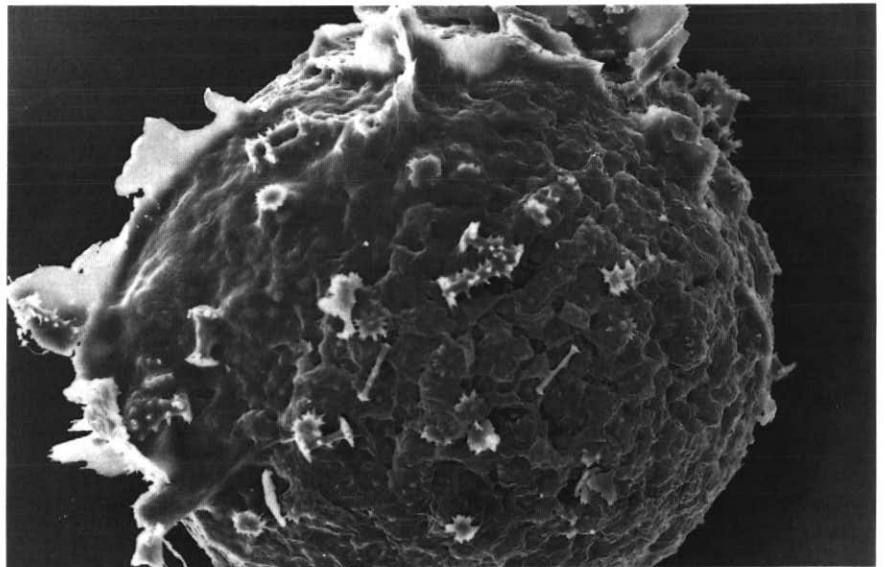
This transition may mark a shift to a more moderate, warm, wet environment. The open boreal woodland was replaced by an open deciduous woodland dominated by *Quercus* and *Ostrya/Carpinus*.

The mid-Holocene interval (7500 to 3500 years B.P.) was a warmer, drier period with low soil moisture. The pollen record indicates an upland mosaic of vegetation with open woodland and prairie patches. The upland deciduous forest was characterized by tree populations of oak, hickory, and chestnut. On mesic slopes surrounding Jackson Pond, forests contained black ash, willow, sweetgum and blackgum.

During late Holocene time (3500 years B.P. to the present), the climate was warm



**Figure 7** Scanning electron micrography of a gemmule of *Anheteromeyenia ryderi*. Close examination of the figure will reveal the two gemmosclere forms characteristic of the species. The shorter, "flat-head" rotule form is also seen in Figure 6. Magnification 440  $\times$ .



**Figure 8** Scanning electron micrograph of a gemmule of *Ephydatia fluviatilis*. Gemmules of this species exhibit only one class of gemmosclere (Figure 5) as opposed to *A. ryderi* (Figures 6 and 7). Magnification 180  $\times$ .



and moist. Willow populations decreased near Jackson Pond. Open grasslands succeeded at the expense of upland woodlands; grasses increased to comprise up to 20% of the upland pollen assemblage. Fossil pollen of *Petalostemon* occurred throughout this interval, indicating the areal expansion of prairie (Wilkins, 1985).

Freshwater sponge spicule analysis was performed on the complete sediment profile from 20,000 years B.P. to the present. The analyses were performed at about 1000-year intervals. During the full-glacial and late-glacial intervals (20,000 to 11,500 years B.P.), the dominant freshwater sponge species in Jackson Pond was *Heteromeyenia latitenta*. Populations of this sponge co-existed with cool-temperate populations of aquatic plants such as naiads and pondweeds (such as *Najas gracillima*, *N. flexilis*, and *Potamogeton spirillus* found together today in the Great Lakes and New England regions). Populations of this cold-water sponge declined in Jackson Pond in the early Holocene and disappeared during the warm, dry mid-Holocene interval. No *H. latitenta* spicules were found after 6000 years B.P. The known distributional range of extant populations of this sponge is limited to the northeastern United States. This is consistent with the relict boreal populations of spruce, persisting until about 6000 years ago in the cool microclimate around Jackson Pond.

A population of the sponge, *Ephydatia fluviatilis* co-existed with *H. latitenta* and has persisted to the present. This species is favoured by water conditions of high conductivity, CaCO<sub>3</sub> hardness, and alkalinity. It can withstand silt-laden, turbid waters. Macrophyte abundance of oospores of marl-forming charophytes confirm conditions of CaCO<sub>3</sub> hardness and alkalinity, particularly between 20,000 and 7000 years B.P. Between 8000 and 7000 years B.P., a third sponge species, *Anheteromeyenia ryderi*, colonized Jackson Pond. This coincided in time with the fossil evidence for the establishment of upland populations of warm-temperate trees such as chestnut and the colonization of lower slopes and wetlands at the margin of Jackson Pond by warm-temperate species of sweetgum, willow, black gum, buttonbush, Virginia willow, and alder. By 7000 years B.P., the bottomland species of willow, buttonbush, Virginia willow and alder formed extensive thickets in the shallow water of the peripheral littoral zone. This complex of swamp forest and shrub thicket probably served as a vegetative screen to filter out mineral sediment eroding from the uplands, thus reducing turbidity levels of water in Jackson Pond. In the middle Holocene, the marginal thicket may have provided an organic substrate for the direct colonization by *A. ryderi*, a sponge species inhibited by high turbidity. Between 7500 and 3500 years ago, the climate was warmer and drier, water tables were lower, and siltation

levels probably decreased. Correspondingly, the turbidity-inhibited populations of *A. ryderi* flourished as the turbidity-tolerant populations of *E. fluviatilis* decreased significantly. Around 3500 years B.P., with a return to the warm and moist climate of the late Holocene, the water level rose in the basin, substantially reducing the area occupied by swamp forest, and reducing the effectiveness of swamp/thicket barrier in filtering out mineral sediment carried from the uplands. *Ephydatia fluviatilis* once again became the dominant species in Jackson Pond. At present, two sponge species inhabit the pond as co-dominants with the introduction of the cosmopolitan species, *Spongilla lacustris*, within the past 500 years, the historic period of European settlement.

The Serendipity and Jackson Pond studies and current studies involving analysis of sediments from northern Ontario indicate that fossilized remains of freshwater sponge spicules and gemmules are well-preserved in most Quaternary wetland and aquatic deposits. Because spicule and gemmule morphologies are species-specific, the most commonly utilized criteria of neosystematics are available to the paleoecologist.

#### Acknowledgements

All scanning electron micrographs are used through the courtesy of Dr. Michael Poirrier of the University of New Orleans. I wish to express my thanks to Mrs. Shirley Weeks for typing the original manuscript. Drs. Hazel and Paul Delcourt, University of Tennessee, Knoxville provided access to their Jackson Pond core and have been an unfailing resource.

#### References

- Delcourt, P.A., 1985, The influence of late-Quaternary climatic and vegetational change on paleohydrology in unglaciated eastern North America: *Ecologia Mediterranea*, v. 11, p. 17-26
- Delcourt, H.R., Delcourt, P.A., Wilkins, G.R. and Smith, E.M., Jr., 1986, Vegetational history of the Cedar Glades regions of Tennessee, Kentucky, and Missouri during the past 30,000 years: *Association of Southeastern Biologists, Bulletin*, v. 33, p. 128-137.
- Frey, D.G., 1964, Remains of animals in Quaternary lake and bog sediments and their interpretation: *Ergebnisse der Limnologie*, v. 2, p. 1-114
- Hall, B.V. and Hermann, S.J., 1980, Paleolimnology of three species of freshwater sponges (Porifera: Spongillidae) from a sediment core of a Colorado semidrainage mountain lake. *American Microscopical Society, Transactions*, v. 99, p. 93-100
- Harrison, F.W., 1974, Sponges (Porifera: Spongillidae), in Hart, C.W. and Fuller, J.L.H., eds., *Pollution Ecology of Freshwater Invertebrates*: Academic Press, New York, p. 29-66.
- Harrison, F.W., 1977, The taxonomic and ecological status of the environmentally restricted spongillid species of North America III *Corvomeyenia carolinensis* Harrison 1971: *Hydrobiologia*, v. 56, p. 187-190.
- Harrison, F.W., 1979, The taxonomic and ecological status of the environmentally restricted spongillid species of North America. V *Ephydatia subtilis* Wetner and *Stratospongilla penneyi* sp. nov.: *Hydrobiologia*, v. 65, p. 99-105
- Harrison, F.W., 1981, Scanning electron microscopy of taxonomic diagnostic criteria of the freshwater sponge, *Heteromeyenia tubisperma* (Potts 1881) (Porifera: Spongillidae): *Hydrobiologia*, v. 77, p. 257-259.
- Harrison, F.W., Gleason, P.J. and Stone, P.A., 1979, Paleolimnology of Lake Okeechobee, Florida: an analysis utilizing spicular components of freshwater sponges (Porifera: Spongillidae): *Notulae Naturae of the Academy of Natural Sciences of Philadelphia*, v. 454, p. 1-6.
- Harrison, F.W. and Harrison, M.B., 1977, The taxonomic and ecological status of the environmentally restricted spongillid species of North America. II *Anheteromeyenia biceps* (Lindenschmidt, 1950): *Hydrobiologia*, v. 55, p. 167-169.
- Harrison, F.W. and Harrison, M.B., 1979, The taxonomic and ecological status of the environmentally restricted spongillid species of North America. IV *Spongilla heterosclerifera* Smith, 1918: *Hydrobiologia*, v. 62, p. 107-111.
- Harrison, F.W., Johnston, L., Stansell, K.B. and McAndrew, W., 1977, The taxonomic and ecological status of the environmentally restricted spongillid species of North America. I *Spongilla spongiosa* Penney, 1957: *Hydrobiologia*, v. 53, p. 99-202.
- Harrison, F.W. and Warner, B.G., 1986, Fossil freshwater sponges (Porifera: Spongillidae) from western Canada: An overlooked group of Quaternary paleoecological indicators: *American Microscopical Society, Transactions*, v. 105, p. 110-120.
- Jewell, M.E., 1935, An ecological study of the freshwater sponges of northeastern Wisconsin *Ecological Monographs*, v. 5, p. 463-504.
- Jewell, M.E., 1939, An ecological study of the freshwater sponges of northeastern Wisconsin, II. The influence of calcium: *Ecology*, v. 20, p. 11-28.
- Old, M.C., 1932, Environmental selection of the freshwater sponges (Spongillidae) of Michigan *American Microscopical Society, Transactions*, v. 51, p. 129-136.
- Pennak, R.W., 1978, *Freshwater Invertebrates of the United States* John Wiley and Sons, New York, 803 p.
- Penney, J.T. and Racek, A.A., 1968, Comprehensive revision of a worldwide collection of freshwater sponges (Porifera: Spongillidae): *Bulletin of the United States National Museum*, v. 272, p. 1-184.
- Poirrier, M.A., 1969, Louisiana freshwater sponges: ecology, taxonomy, and distribution, unpublished Ph.D. Thesis, Louisiana State University, 172 p.
- Poirrier, M.A., 1974, Ecomorphic variation in gemmoscleres of *Ephydatia fluviatilis* Linnaeus (Porifera: Spongillidae) with comments upon its systematics and ecology: *Hydrobiologia*, v. 44, p. 337-347
- Poirrier, M.A., 1977, Systematic and ecological status of *Anheteromeyenia ryderi* (Porifera: Spongillidae) in Louisiana: *American Microscopical Society, Transactions*, v. 96, p. 62-67.
- Racek, A.A., 1966, Spicular remains of freshwater sponges: *Memoirs of the Connecticut Academy of Arts and Sciences*, v. 17, p. 78-83.

- Racek, A.A., 1970, The Porifera, in Hutchinson, G.E., ed., *Ianula: An account of the history and development of the Lago di Monterosi, Latium, Italy*: American Philosophical Society, Transactions, v. 60, p. 143-149.
- Racek, A.A., 1974, The waters of Merom: A study of Lake Huleh. IV. Spicular remains of freshwater sponges (Porifera): *Archiv für Hydrobiologie*, v. 74, p. 137-158.
- Racek, A.A. and Harrison, F.W., 1975, The systematic and phylogenetic position of *Paleospongilla chubutensis* (Porifera: Spongillidae): *Linnean Society of New South Wales, Proceedings*, v. 99, p. 157-165.
- Simpson, T.L., 1984, *The Cell Biology of Sponges*: Springer-Verlag, New York, 661 p.
- Turner, J., 1985, Sponge gemmules from lake sediments in the Puget Lowland, Washington: *Quaternary Research*, v. 24, p. 240-243.
- Warner, B.G., 1984, Late Quaternary paleoecology of eastern Graham Island, Queen Charlotte Islands, British Columbia, Canada, unpublished Ph.D. Thesis, Simon Fraser University, British Columbia, 190 p.
- Wilkins, G.R., 1985, Late-Quaternary vegetational history at Jackson Pond, Larue County, Kentucky, unpublished M.Sc. Thesis, University of Tennessee, Knoxville, 172 p.

Accepted 29 December 1987.



From the **GEOLOGICAL ASSOCIATION OF CANADA**

A collection of reviews originally published in *Geoscience Canada*.

Topics include:

- uranium series disequilibrium dating
- tephrochronology and fission-track dating
- the promise of atom counting
- amino acid racemization
- paleomagnetism
- weathering
- thermoluminescence
- electron spin resonance

**Ideal as a university text!** 1985, \$12.00, 87 pages, paper

Order from:

Geological Association of Canada  
Association géologique du Canada

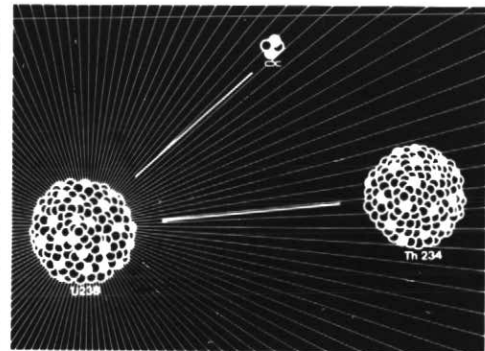
Department of Earth Sciences, 4 Clark Place, Memorial University  
of Newfoundland, St. John's, Newfoundland A1B 3X5

## DATING METHODS OF PLEISTOCENE DEPOSITS AND THEIR PROBLEMS

**Geoscience  
canada**

Reprint Series 2

Edited by  
N.W. Rutter



**VISA or MASTERCARD** now accepted. Please add \$3.00 per book for postage and handling. Payment by cheque or money order may also be made in equivalent U.S. funds. Make cheque payable to **Geological Association of Canada**. If paying by VISA or MASTERCARD send full card number, expiry date and signature. Payment must accompany orders.